



CORPUS PUBLISHERS

Corpus Journal of Dairy and Veterinary Science (CJDVS)

Volume 1 Issue 1, 2020

Article Information

Received date: May 15, 2020

Published date: May 23, 2020

*Corresponding author

Márcio Garcia Ribeiro, Department of Animal Production and Preventive Veterinary Medicine, UNESP, Botucatu, SP, Brazil

Distributed under Creative Commons CC-BY 4.0

Keywords Mammary protothecosis; Protothecal bovine mastitis; *In vitro* susceptibility pattern

Mini Review

Prototheca-Induced Infections in Dairy Cows: A Multiresistant Environmental Pathogen

Márcio Garcia Ribeiro*

School of Veterinary Medicine and Animal Sciences, São Paulo State University-UNESP, Botucatu, Brazil

Abstract

Prototheca spp is an achlorophyllous alga found ubiquitously in the environment. The organism is opportunistic in nature and has been considered an emerging disease that affects humans and some domestic animals, although bovine mastitis is a major clinical manifestation of algae-infections among livestock. The pathogen is commonly nonresponsive to conventional therapy, which has led to segregation and premature culling of chronic cases as main measures to control of protothecal mastitis. In this scenario, this mini-review focused on epidemiological aspects, *in vitro* resistance of algae, and measures applied to control and prevention of protothecal mammary infections in dairy cows.

Etiology

Members of the genus *Prototheca* consist of unicellular, colorless, achlorophyllous intracellular yeast-like micro-algae able to infect humans [1] and animals [2]. Based on a current taxonomic proposal using mitochondrial *cytb* gene molecular-marker, fourteen *Prototheca* species are assigned: typical dairy cattle-associated, i.e., *Prototheca bovis* (formerly *Prototheca zopfii* genotype 2), *Prototheca blaschkeae*, and *Prototheca ciferrii* (formerly *Prototheca zopfii* genotype 1); human-associated, i.e., *Prototheca wickerhamii*, *Prototheca cutis*, and *Prototheca miyajii*, as well as *Prototheca moriformis* (*Prototheca ulmea*), *Prototheca stagnora*, *Prototheca tumulicola*, *Prototheca zopfii*, *Prototheca cookei*, *Prototheca pringsheimii*, *Prototheca xanthoriae*, and *Prototheca cerasi*, the last four ones, recently named [3]. Among these species, *Prototheca ciferrii* has been found in manure and environment of dairy farms, whereas *Prototheca blaschkeae* and *Prototheca bovis*, have been described globally causing mammary infections, besides predominance of *Prototheca bovis* as a most frequent cause of clinical protothecal mastitis in dairy herds [2].

Epidemiology

Prototheca species inhabit a wide variety of environmental niches and surfaces, particularly with high humidity and abundant organic matter contents [1,2]. On endemic-dairy farms, these saprophytic algae have been recovered in farm-sources where animals are driven or resting, including streams or stagnant ponds, mud and manure surround milking areas, feed, cattle drink water, feces of cattle and calves, the material of cow's bed and compost barn, as well as milking machine [4,5]. Improper teat hygiene procedures and poor hygiene in the intramammary infusions approach, particularly the use of dry cow teat sealant and infusions with nonintramammary formulations, revealed strong herd-risk factors to protothecal mastitis [6]. Transmission between infected-to-healthy hosts may also occur due to deficiencies in milking hygiene procedures, improper pre-milking practices, and excess of organic material on the environment of dairy cows [4]. Variable prevalence (<10%) of *Prototheca* species isolation has been seen among mastitis cases and bulk tank milk samples subjected to microbiological culture worldwide [2,6-8]. Nonetheless, increasing rates of clinical protothecal mastitis have been reported globally [2,3,5], with marked outbreaks recorded in South (Brazil) and North America (USA, Canada, and Mexico), Asia (Japan and China), Oceania (New Zealand), and some countries of Europe (Denmark, England, and Italy). A large-scale study involving molecular characterization of 342 *Prototheca* isolates obtained of bovine mastitis from different countries confirmed high prevalence of *Prototheca bovis* (90.6%), followed by minor frequency of *Prototheca blaschkeae* (8.8%), and *Prototheca ciferrii* (0.6%), reinforcing *Prototheca bovis* as a major cause of mammary infections in cattle-herds [2].

Pathogenicity and Clinical Aspects

The virulence and pathogenicity of *Prototheca*-induced infections remain poorly understood, although the rigid structure of cell wall, suppression of cell-mediated immune response, evasion of phagocytic cells [1,9], and biofilm-formation [10,11] probably enables intracellular persistence that, in turn, results in pyogranulomatous reactions with the destruction of mammary tissues and epithelia, leading to limited immune response and tissue resolution [9]. Mostly cows develop clinical indurative mastitis, during lactation or dry period. Mammary protothecosis produces a decrease in milk yield, as opposed to increasing SCC. Sporadically the pathogen spread from mammary glands to infect other organs. Some animals may develop subclinical form or solve clinical signs spontaneously, although the pathogen tends long-term infections [4]. Mammary infections usually are refractory to conventional intramammary and/or systemic therapy [12], which results in poor prognosis or outcome, especially when two or more teats are affected. In this regard, on endemic-farms, economic losses have been related to the deep impact of decrease milk production and early culling of clinical cases [2-4,12].

Diagnostic Approach

Routine identification of the *Prototheca* species has been based on conventional microbiological culture, staining (Gram, Romanowsky variants, lactophenol cotton blue), micromorphology, and biochemical activity (carbohydrate and alcohol assimilation) [1,2,5,9]. However, the algae may be eliminated intermittently by milk, a fact that may limit microbiological diagnosis on farms [4]. In the coming years, the speciation, genotyping, taxonomic reclassification, and novel species characterization of algae has been allowed using various molecular methods, e.g., conventional and multiplex PCR, PCR-restriction enzyme analysis (PCR-REA) assay (partial *cytb* gene), sequencing (18S rRNA gene), and MALDI-TOF MS [2,3,6,10].

Treatment

Since the treatment of mammary infections usually is unsuccessful, *Prototheca* sp isolated from milk of dairy cows have been



subjected to a number of *in vitro* studies with a wide variety of pharmacological products with well-known microbicide effect, and have shown susceptibility to some antifungal (amphotericin B, itraconazole, ketoconazole) [13], antimicrobials (gentamicin, netilmicin, colistin sulfate, kanamycin) [11,13] and, recently, dinitroanilines, a group of herbicides [11]. Also, *in vitro* algicidal effect in low concentrations has been seen in antiseptic, disinfectants and/or sanitizing such as iodine, sodium hypochlorite [14], peracetic acid [10], guanidine [15], as well as other products that have been assessed, e.g., hydrogen peroxide, thimerosal, chlorhexidine, copper sulfate, and silver nitrate. Nonetheless, there is no strong association between *in vitro* susceptibility and *in vivo* microbiological cure of mammary infections, in addition to none standard guidelines to *in vitro* susceptibility tests for *Prototheca*. Besides *in vitro* action of some these pharmacological products against *Prototheca* species, commonly have been seen only temporary *in vivo* clinical regression, which no effectively alter the outcome of clinical mammary infections [12] and, then, in a great number of animals, if not all, usually is observed the return of clinical signs.

Control and Prevention

Measures recommended avoiding protothecal mastitis are similar to prevent and control other environmental pathogens, due to opportunistic behavior and environmental nature of the algae [4]. Therefore, these cares include a routine of clinical/subclinical and microbiological diagnosis of mastitis, adequate milking hygiene, teat dip practices, offering food after milking, water chlorination used in milking procedures, clean and dry pre- and postmilking area, and management of bedding and compost barn material. On endemic-herds, early diagnosis, segregation of affected animals to end of milking, dry off teat (only one teat affected), or culling of multi-teat and/or chronic animals appears to be main measures to control protothecal mammary infections.

Public Health Issue

Ingestion of contaminated milk and derivatives from bovines has been seen as a source of the transmission of the *Prototheca* species from cows-to-people [16], and it poses as a public health concern because these algae may resist to industrial heat temperature applied to milk and milk products intended to human consumption, and cause gastroenteritis signs; although cutaneous lesions, articular disorders, and systemic or disseminated infections have also been reported in human protothecosis.

References

1. Lass Flörl C, Mayr A (2007) Human protothecosis. *Clinical Microbiology Reviews* 20: 230-242.
2. Ahrholdt J, Murugaiyan J, Straubinger RK, Jagielski T, Roesler U (2012) Epidemiological analysis of worldwide bovine, canine and human clinical *Prototheca* isolates by PCR genotyping and MALDI-TOF mass spectrometry proteomic phenotyping. *Medical Mycology* 50(3): 234-243.
3. Jagielski T, Bakula Z, Gawor J, Maciszewski K, Kusber W, et al. (2019) The genus *Prototheca* (Trebouxiophyceae, Chlorophyta) revisited: Implications from molecular taxonomic studies. *Algal Research* 43: 101639.
4. Ruegg PL (2016) Mastitis in large animals. *The Merck Veterinary Manual* (11th edn), Merck & Co., Inc. Kenilworth, NJ, USA, pp. 1358-1368.
5. Kano R (2019) Emergence of fungal-like organisms: *Prototheca*. *Mycopathologia*.
6. Pieper L, Godkin A, Roesler U, Polleichtner A, Slavic D, et al. (2012) Herd characteristics and cow-level factors associated with *Prototheca* mastitis on dairy farms in Ontario, Canada. *Journal of Dairy Science* 95(10): 5635-5644.
7. Park HS, Moon DC, Hyun BH, Lim SK (2019) Short communication: Occurrence and persistence of *Prototheca zopfii* in dairy herds of Korea. *Journal of Dairy Science* 102(3): 2539-2543.
8. Jagielski T, Roeske K, Bakula Z, Piech T, Wlazlo L, et al. (2019) A survey on the incidence of *Prototheca* mastitis in dairy herds in Lublin province, Poland. *Journal of Dairy Science* 102(1): 619-628
9. Quinn PJ, Markey BK, Leonard FC, Fitzpatrick ES, Fanning S, et al. (2011) Pathogenic algae and cyanobacteria. *Veterinary Microbiology and Microbial Disease* (2nd edn), Wiley-Blackwell, West Sussex, UK, pp. 478-482.
10. Gonçalves JL, Lee SHIN, Arruda EP, Galles DP, Caetano VC, et al. (2015) Biofilm-producing ability and efficiency of sanitizing agents against *Prototheca zopfii* isolates from bovine subclinical mastitis. *Journal of Dairy Science* 98(6): 3613-3362.
11. Morandi S, Cremonesi P, Capra E, Silvetti T, Decimo M, et al. (2016) Molecular typing and differences in biofilm formation and antibiotic susceptibilities among *Prototheca* strains isolated in Italy and Brazil. *Journal of Dairy Science* 99(8): 6436-6445.
12. Morello L, Tommaso T, Aretino F, Morandi S, Breviaro D (2020) Preliminary results, perspectives, and proposal for a screening method of *in vitro* susceptibility of *Prototheca* species to antimicrotubular agents. *Antimicrobial Agents and Chemotherapy* 64: e01392-19.
13. Jagielski T, Buzzini P, Lassa H, Malinowski E, Branda E, et al. (2012) Multicentre Etest evaluation of *in vitro* activity of conventional antifungal drugs against European bovine mastitis *Prototheca* spp. isolates. *Journal of Antimicrobial Therapy* 67(8): 1945-1947.
14. Salerno T, Ribeiro MG, Langoni H, Siqueira AK, Costa EO, et al. (2010) *In vitro* algacide effect of sodium hypochlorite and iodine based antiseptics on *Prototheca zopfii* strains isolated from bovine milk. *Research in Veterinary Science* 88(2): 211-213.
15. Alves AC, Capra E, Morandi S, Cremonesi P, Pantoja JCF, et al. (2017) *In vitro* algicidal effect of guanidine on *Prototheca zopfii* genotype 2 strains isolated from clinical and subclinical bovine mastitis. *Letters in Applied Microbiology* 64(6): 419-423.
16. Constable PD, Hinchliff KW, Done S, Gruenberg W (2016) Diseases of the mammary gland. *Veterinary medicine: A textbook of the diseases of cattle, horses, sheep, pigs, and goats*. (11th edn) Saunders Ltd., Philadelphia, USA pp. 1904-1996.